



LIFE MEDICAL DIAGNOSTICS – Abu Dhabi

TEST COLLECTION GUIDE 2022



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CONTACT INFORMATION

Contact Details

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Operation Time:
Sunday - Saturday: 8:00 AM to 11:00 PM

To continue to improve and provide better service, we need the valuable feedback and suggestions from you, as our valued client. If you have a complaint or feedback (positive/negative) and for any unanswered questions regarding specimen requirements, test availability, and specimen handling arise, please feel free to contact us.

The staff of Life Medical Diagnostics is committed to providing you with accurate information and courteous service. We will strive to exceed your expectations and provide you with quality assured medical service.





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OUR MISSION

Life Medical Diagnostic Centre is committed to providing focused, high quality, cost-effective laboratory services to our patients and physicians with the continuous quality improvement which enhances patient health in a safe and supportive environment.

OUR VISION

To provide the highest quality, state-of-the-art- laboratory testing and expert pathology diagnosis to our patients in an affordable and timely manner.

OUR VALUES

- We are committed to trust, respect and integrity.
- We promote and facilitate excellence and Innovation.
- We always provide service with compassionate care.
- We approach everything through team work and continuous learning and education.



SPECIMEN REQUIREMENTS

- **Identification and Test Request Forms**

Complete a test request form for each patient. Do not fill in more than one form per patient. Please make sure that the age and gender are included.

- **Clinical Information**

The clinical information form should be attached to the request form.

- **Tube Labeling**

Label each sample tube with at least two identifiers: patient's full name and file number.

- **Storage and Transportation**

The quality of the results produced relies heavily on the quality of the specimen received. Samples must be maintained at the required temperature all the times.

- **Turnaround Time (TAT)**

All routine tests are performed as arrived to the laboratory. However, few tests are batches and run on scheduled days.

The Laboratory Management ensures that all patients are getting appropriate care in a timely manner. For complete TAT refer to the test menu.

Please note that turnaround time (TAT) in this Test Directory only applies to tests performed in Life Medical Diagnostics – Abu Dhabi. For exact TAT of other tests not included in the list, please contact our Customer Care Department.

COAGULATION TESTS SAMPLE PREPARATION

1. Establish the identity of the patient.
2. Draw blood into a light blue-topped tube that contains 3.2% (0.109 M) sodium citrate anticoagulant. Allow the tube to fill to the proper level; a short draw critically affects the result as there is excess anticoagulant relative to blood volume.
3. Gently invert six times to mix.
4. Label the tube with patient's full name, medical record number, and the date and time of collection.
5. Centrifuge the stoppered tube at 2500 g for 15 minutes.
6. Transfer the plasma with a pipette into a plastic tube. Cap the tube and centrifuge it for additional 15 minutes at 2500 g.
7. Taking care not to disturb the cell button at the bottom of the tube, transfer the plasma to another clean plastic tube. Label the tube with patient's full name, medical record number, the date and time of collection and write "Citrate Plasma".
8. When plasma testing cannot be completed within 4 hours of collection, freeze the plasma immediately.
9. A completed request form with a brief patient history must accompany each specimen. Anticoagulant such as heparin, warfarin and others may affect many test results and must be noted on the test request form.

SPECIMEN TRANSPORTATION ROUTINE SPECIMEN

1. Place the specimen or the specimens of the same type from one patient in the sample compartment of the biohazard bag and seal the zip. The laboratory request should be placed in the outside pocket of the biohazard bag. Place the appropriate accompanying order forms, as applicable, together with the laboratory request.

NB: One bag for one specimen type for one patient only.

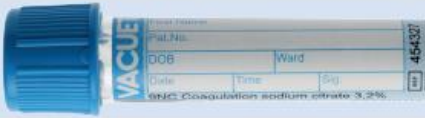


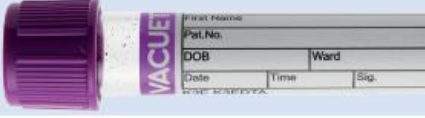

2. For patient with single laboratory request requiring multiple specimen types, attach the biohazard bags together with the request.
3. Place other specimen types, such as urine, stool, or microbiology swabs into a separate biohazard bag.
4. To prevent leakage, seal containers with parafilm.
5. Container for 24 – hour urine specimen must be sealed tightly and placed in a large biohazard bag. Write the total volume of the urine in the laboratory request form.
6. Container for specimens that must be protected from light must be wrapped in aluminum foil and place in a biohazard bag.
7. Do not refer specimens without approved biohazard packaging. Our pre – analytical unit staff have instructions to reject specimens that arrive without biohazard packaging. Leaking specimens are rejected as well.

PHLEBOTOMY PROCEDURE (CLOSED SYSTEM)

1. Identify the patient. Confirm patient identification by asking an open ended question like “What is your name?” **NEVER DRAW IF THERE IS A DISCREPANCY**
2. Identify yourself and explain the procedure.
3. Assess the patient's physical disposition (i.e. diet, exercise, stress, basal state). Verify fasting states and/or diet restrictions.
4. Check the requisition form for requested tests, patient information, and any special requirements.
5. Ask the patient if he/she has a latex sensitivity for the used gloves.
6. Perform hand hygiene and then wear gloves.
7. Assemble your supplies.
8. Apply the tourniquet and select a suitable site for venipuncture. Make sure not to apply the tourniquet for more than one minute.
9. Release tourniquet. Cleanse the venipuncture site by using 70% isopropyl alcohol. Allow to air dry for at least 30 seconds.
If you MUST re-touch, then you MUST re-CLEAN!
10. Reapply tourniquet. Ask the patient to make a fist without pumping the hand. Perform venipuncture. Once blood flow begins, request patient to open hand.
11. **Use the correct order of draw according to the CLSI guidelines.** Invert tubes after filling. **Do a full draw especially for tubes with anticoagulant.**
12. When the last tube to be drawn is filling, release and remove the tourniquet.
13. Place the gauze pad over the puncture site and then remove the needle using a swift backward motion.
14. Press down on the gauze applying adequate pressure making sure bleeding has stopped, and apply the adhesive bandage.
15. Recognize complications associated with the phlebotomy procedure.
16. Assess the need for sample recollection and/or rejection.
17. **Label the specimen containers** with at least two identifiers: patient's full name and file number **in the presence of the patient after extraction.**
18. Dispose of contaminated materials/supplies in designated containers.

Order of Draw

based on CLSI GP41-ED7

1	Blood Culture	
2	Sodium Citrate/CTAD*	
3	Serum with and without Separator	
4	Heparin with and without Separator	
5	EDTA with and without Separator	
6	Glycolytic Inhibitor	
7	Other additives	

* If a winged blood collection set is used, the first tube in the series will be under-filled. Therefore, if a Sodium Citrate specimen is drawn first, a discard tube (No Additive) is recommended to be drawn prior to this tube to ensure the proper additive-to-blood ratio. In addition, even though studies have shown that PT and aPTT tests are not affected if drawn first in a tube series, it is advisable to draw a second tube for other coagulation assays, since it is not known whether or not these tests will be affected.

NOTE: Always follow your facility's protocol for order of draw.

ADVERSE REACTIONS DURING SPECIMEN COLLECTION

At any time a patient will show adverse reaction during specimen collection, the procedure shall be immediately stopped.

All adverse reactions post phlebotomy procedure must be reported to the authorized personnel.

ADVERSE REACTION	ACTION TO BE TAKEN
Nausea and Vomiting	Terminate the procedure. Ask the patient to breath slowly and deeply. Apply a cold compress to the patient's forehead and provide the patient with an emesis basin.
Fainting or Syncope (State of loss of consciousness and postural tone resulting from insufficient blood flow to the brain)	Watch the patient for signs of fainting like pallor (paleness), perspiration, hyperventilation, vertigo, dizziness, and light-headedness. If fainting occurs, discontinue immediately the procedure. Have the patient to lie down on his/her back, raise their foot above the level of their head, loosen tight clothing, apply a cold compress to their forehead, and check for their pulse. If the patient is not responding, seek medical help. When the patient regains consciousness, the patient must remain in the area for at least 15 minutes and then should be instructed not to operate any vehicle for at least 30 minutes.
Pain	Assure the patient to help them relax and then the procedure will be less painful. Avoid any excessive, deep, blind or lateral redirection of the needle. When patient feels pain in his/her arm, immediately discontinue the procedure. Ask the patient if the pain has stopped. If so, try to redraw at another site if the patient is willing. Application of an ice pack to the site after needle removal can help prevent or reduce inflammation associated with nerve involvement.
Petechiae (Tiny, non-raised red spots that appear on the patient's skin when a tourniquet is applied)	The presence of petechiae indicates that venipuncture site may bleed excessively. Therefore, the procedure shall be immediately terminated.
Seizures/Convulsion	Terminate the procedure. Ensure that the patient doesn't possess anything in his mouth. Seek immediate help from any nearby or other phlebotomist and the physician.
Hematoma Formation	Discontinue the draw and the specimen shall then be discarded since it may become hemolyzed and can still produce inaccurate results. Apply a pressure over the puncture site for at least two minutes. A cold compress or an ice pack must also be applied on the puncture site to relieve pain and reduce swelling.
Allergy	If the patient starts to feel a difficulty in breathing, the phlebotomist must rapidly stop the procedure. If an anaphylactic shock reaction takes place and the patient completely stops breathing, immediately perform CPR and ask for faster help and support.

ADVERSE REACTIONS DURING SPECIMEN COLLECTION (continued)



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ADVERSE REACTION	ACTION TO BE TAKEN
<p>Phlebitis (characterized by the presence of a tender red area on the skin)</p>	<p>Discontinue the procedure immediately.</p>
<p>Vasovagal Reaction</p>	<p>Discontinue the procedure immediately and help control the patient to prevent injury. Immediately call for help from trained medical staff.</p>
<p>Anxiety/Fear</p>	<p>Start relieving the patients by explaining exactly what will happen and tries to calm him down as much as possible. Communicate with the patient on each step he is doing to keep on reassuring him.</p>
<p>Shock</p>	<p>Maintain an open airway for the victim by keeping him lying down with the head lower than the rest of the body. Keep the patient warm and to call for immediate assistance.</p>
<p>Excessive Bleeding</p>	<p>Apply a firm pressure on the puncture site until the bleeding visibly ceases. Advise the patient to hold his hand elevated above the level of his/her heart.</p>

GENERAL INSTRUCTIONS FOR MICROBIOLOGY SPECIMEN COLLECTION AND HANDLING

Specimen collection:

1. Make every effort to obtain specimens prior to the initiation of antimicrobial therapy.
2. Use strict aseptic technique.
3. Obtain specimens from normally sterile sites, taking care to minimize contamination by the normal colonizing flora of the skin or mucous membranes.
4. Collect an adequate volume of specimen; send tissue or fluid whenever possible rather than submitting a specimen collected on a swab.
5. If the specimen is collected in a syringe, remove the needle and replace it with a cap prior to transport.
6. Label all specimen containers with identifying information about the patient (Full patient name and file number), the specimen source and date and time of collection.
7. Fill out all requisitions completely and precisely, including requested details on patient history, antimicrobial therapy, and specimen source, so that the laboratory can best determine the appropriate method for processing the specimen.
8. Notify the laboratory in advance if special tests are requested or if unusual pathogens, including potential agents of bioterrorism, are suspected.
9. Place warning labels on specimens from patients suspected of having highly contagious diseases and notify the laboratory.

Specimen transportation:

1. In general, specimens may be sent to the microbiology laboratory in sterile, leak proof containers enclosed within leak proof, sealed biohazard plastic bags.
2. Prompt transport of specimens to the microbiology laboratory is essential in order to optimize the yield of cultures and the interpretation of results (CSF, wound cultures, anaerobes). Delays in processing may result in the overgrowth of some microorganisms or the death of more fastidious ones.
3. Desiccation of the sample must also be avoided.
4. Use of transport media when necessary and methods for preservation of specimens if processing is delayed (e.g., refrigeration of urines).
5. Specimens for mycobacterial culture should be collected appropriately and transported to the laboratory without delay.
6. The laboratory recommend collecting 3 sputum specimens for acid-fast smears and culture in patients with clinical and chest x-ray findings compatible with tuberculosis. Specimens must be delivered to the laboratory promptly, ideally within 30 minutes, but at least within 1 day of collection. Specimens should be refrigerated within 1 hour of collection.
7. All specimens for mycobacterial culture are collected and/or received in sealed leak-proof containers.

BLOOD CULTURE SPECIMEN COLLECTION

Blood culture specimens to our lab & to eliminate the high blood culture contamination rate, please the following pre analytic instructions is very important to be followed in order to decrease this high rate:

Patient Preparation and Blood Collection for Blood Culture

Timing of blood collection:

1. Whenever possible, blood should be taken before antibiotics are administered.
2. The best time is when the patient is expected to have chills or a temperature spike.
3. It is recommended that two or preferably three blood culture be obtained, separated by intervals of approximately one hour or less if treatment cannot be delayed or within 24-hours period. More than three blood cultures are rarely indicated.
4. The advantages of repeated cultures are as follows:
 - The chance of missing a transient bacteraemia is reduced.
 - The pathogenic role of “saprophytic” isolates (e.g. *Staphylococcus epidermidis*) is confirmed if they are recovered from multiple venipuncture.

Skin Disinfection:

1. The skin at the vein puncture site must be meticulously disinfected using 70% isopropyl alcohol followed by 2% tincture of iodine, 10% polyvidone iodine, 70% alcohol, or 0.5% chlorhexidine in 70% alcohol.
2. The alcohol and disinfectant should be allowed to evaporate on the skin surface before blood is withdrawn. If tincture of iodine is used, it should be wiped off with 70% alcohol to avoid possible skin irritation.

Recommended Volume:

1. For Adult: 20-30 ml blood per venipuncture distributed equally between aerobic and anaerobic bottles.
2. For Children and Infants: it should not be more than 1% of total patient blood volume per venipuncture.

Filling the Vials:

1. Before filling the vials, the rubber plug should be disinfected using 70% isopropyl alcohol left to dry.
2. The aerobic culture bottle must be filled before the anaerobic culture bottle.

There is no need to change syringe needle between the two vials. Bottles should be inverted gently for several times to inhibit blood clotting.

GENERAL INSTRUCTIONS FOR NASOPHARYNGEAL SWAB COLLECTION FOR COVID-19 BY PCR

Specimen collection:

1. Wear your full PPE's probably, and adequately.
 - Full body isolated apron with overhead cover.
 - Double Gloves
 - N95 Mask
 - Goggles
 - Face shield
2. Use only synthetic fiber swabs with plastic shafts. Do not use calcium alginate swabs or swabs with wooden shafts or cotton swabs; as they may contain substances that can inactivate some viruses and inhibit PCR testing.
3. Insert a swab into the nostril parallel to the palate. Leave the swab in place for a few seconds to absorb secretions. Swab both nasopharyngeal areas with the same swab:
 - Pass the nasopharyngeal swab through the nares.
 - Resistance will be met, and this will confirm contact with the nasopharynx.
 - Rub the swab tip several times across the mucosal surface. This will loosen and collect cellular material.
 - Allow a time of contact of up to 30 seconds.
 - Withdraw the swab and place it into the viral transport media (VTM) make sure liquid medium covers the swab tip.
 - Break or cut the end of the swab, and screw vial lid on tightly.
 - Properly label the sample.

Specimen transportation:

1. Clients should fill out the regular request form of with patient data.
2. Refrigerate swabs at 2-8 °C until shipping or testing.
3. Life Medical Diagnostics uses collection boxes; with gel ice bag for cooling and maintaining temperature at 2-8 °C.

GENERAL CRITERIA FOR REJECTION/PENDING OF SPECIMEN

Unacceptable specimens may result in test cancellation or delay. Listed below are general causes of such action.

A. Rejected Samples

1. Unlabeled specimen (by file no. and patient name).
2. Discrepancy between request and sample label.
3. Wrong/Inappropriate transportation condition.
4. Insufficient sample volume.
5. Wrong sample (collected on a wrong tube).
6. Complete leaking of the sample.
7. Under- or overfilled collection tubes.
8. Grossly hemolyzed samples.
9. Grossly lipemic samples.
10. Urine trace metal not in acid washed tube.
11. Samples that must be frozen received refrigerated.
12. Samples that must be protected from light not wrapped with aluminum foil. Examples of tests that are needed to be protected from light are:
 - Bilirubin
 - Porphobilinogen
 - Porphyrines
 - Vitamin B1, B2, and B6
 - Carotene
13. Unfixed Pathology and cytology specimen or improper fixation.
14. Broken Histopathology slides.

B. Pending Samples

1. No or insufficient bio data (e.g. Incomplete name, no age...)
2. No report if required by the test being requested.
3. Pathology or Cytology specimens w/o report

SPECIFIC MICROBIOLOGY SPECIMEN REJECTION CRITERIA

Aerobic Culture

1. Body fluids received in swabs.
2. Dried swabs & Improper transport media.
3. Clotted blood culture samples or containers with anticoagulant.
4. Gram stain for Neisseria gonorrhoea on specimens from cervix, vagina, and anal crypts.
5. Specimen received in unsterile container or grossly contaminated
6. Foley catheter tip.
7. Pooled 24 hour urine or sputum received for culture.
8. Unsatisfactory expectorated sputum (saliva in place of sputum).
9. Duplicate specimen (other than duplicate blood)
10. Intrauterine Device (IUD) Inappropriate for culture as contaminated with normal vaginal flora.
11. Specimens submitted in a collection device that has not been used to validate the procedure.
12. Specimen types that have not been validated for the procedure.
13. Stool specimens for bacterial culture are not accepted from inpatients after the third hospital day and no more than two stool specimens /per patient without prior consultation.
14. Tissue specimen for cultures submitted in formalin.

Anaerobes Culture

1. Specimen for anaerobes not received in appropriate container.
2. Anaerobic culture requested on improper specimen.



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GENERAL TEST MENU

Tests marked with ****** are under ISO 15189:2012 Scope of Accreditation.

A/G RATIO (ALBUMIN/GLOBULIN RATIO)

CODE	
SPECIMEN	1 mL Serum
TEMPERATURE	2 – 8°C
SETUP SCHEDULE	DAILY
TAT	1 day after setup schedule.
METHODOLOGY	Calculated
PATIENT PREPARATION	Not Applicable
COMMENT/S	

Adeno / Rota virus Ag*

CODE	1014
SPECIMEN	Fresh Stool Sample (No transport media)
TEMPERATURE	2 – 8°C
SETUP SCHEDULE	DAILY
TAT	1 day after setup schedule.
METHODOLOGY	Immuno Chromatographic
PATIENT PREPARATION	Not Applicable
COMMENT/S	

ALANINE AMINOTRANSFERASE (ALT)*

CODE	1018
SPECIMEN	1 mL Serum
TEMPERATURE	2 – 8°C
SETUP SCHEDULE	DAILY
TAT	1 day after setup schedule.
METHODOLOGY	IFCC
PATIENT PREPARATION	Not Applicable
COMMENT/S	

ALBUMIN / CREATININE RATIO

SEE Microalbuminuria (Albumin / Creatinine Ratio) for details.

ALBUMIN*

CODE	1020
SPECIMEN	1 mL Serum
TEMPERATURE	2 – 8°C
SETUP SCHEDULE	DAILY
TAT	1 day after setup schedule.
METHODOLOGY	BCG

PATIENT PREPARATION Not Applicable
 COMMENT/S

ALBUMIN IN 24 HOUR URINE

CODE	1022
SPECIMEN	5 mL of 24 hour urine sample
TEMPERATURE	2 – 8°C
SETUP SCHEDULE	DAILY
TAT	1 day after setup schedule.
METHODOLOGY	Turbidimetric
PATIENT PREPARATION	At early morning discard the first sample of urine and then record the time on the container. Collect all urine specimens in a bottle for the next 24 hours until the next morning at the same time of the first collected sample the day before. Keep the container refrigerated during the whole process.
COMMENT/S	Total urine volume is required.

ALKALINE PHOSPHATASE*

CODE	1026
SPECIMEN	1 mL Serum
TEMPERATURE	2 – 8°C
SETUP SCHEDULE	DAILY
TAT	1 day after setup schedule.
METHODOLOGY	IFCC
PATIENT PREPARATION	Not Applicable
COMMENT/S	

ALPHA FETO PROTEIN (AFP)*

CODE	1031
SPECIMEN	1 mL Serum
TEMPERATURE	2 – 8°C
SETUP SCHEDULE	AS NEEDED
TAT	1 day after setup schedule.
METHODOLOGY	Chemiluminescence Immunoassay
PATIENT PREPARATION	Not Applicable
COMMENT/S	

AMYLASE IN SERUM*

CODE	1043
SPECIMEN	1 mL Serum
TEMPERATURE	2 – 8°C
SETUP SCHEDULE	DAILY
TAT	1 day after setup schedule.
METHODOLOGY	IFCC
PATIENT PREPARATION	Not Applicable
COMMENT/S	



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Anti CCP Abs

CODE	1055
SPECIMEN	1 mL Serum
TEMPERATURE	2 – 8°C
SETUP SCHEDULE	AS NEEDED
TAT	1 day after setup schedule.
METHODOLOGY	Chemiluminescence Immunoassay
PATIENT PREPARATION	Not Applicable
COMMENT/S	

Anti ds-DNA Ab*

CODE	1057
SPECIMEN	1 mL Serum
TEMPERATURE	2 – 8°C
SETUP SCHEDULE	DAILY
TAT	1 day after setup schedule.
METHODOLOGY	ELISA
PATIENT PREPARATION	Not Applicable
COMMENT/S	

ANTI MULLERIAN HORMONE (AMH)*

CODE	1071
SPECIMEN	1 mL Serum
TEMPERATURE	2 – 8°C
SETUP SCHEDULE	AS NEEDED
TAT	1 day after setup schedule.
METHODOLOGY	Chemiluminescence Immunoassay
PATIENT PREPARATION	Not Applicable
COMMENT/S	

ANTI THYROGLOBULIN AB*

CODE	1088
SPECIMEN	1 mL Serum
TEMPERATURE	2 – 8°C
SETUP SCHEDULE	AS NEEDED
TAT	1 day after setup schedule.
METHODOLOGY	Chemiluminescence Immunoassay
PATIENT PREPARATION	Not Applicable
COMMENT/S	

ANTI THYROID PEROXIDASE ABS (TPO)*

CODE	1090
SPECIMEN	1 mL Serum

TEMPERATURE	2 – 8°C
SETUP SCHEDULE	AS NEEDED
TAT	1 day after setup schedule.
METHODOLOGY	Chemiluminescence Immunoassay
PATIENT PREPARATION	Not Applicable
COMMENT/S	

ANTISTREPTOLYSIN O TITER (ASOT)*

CODE	1096
SPECIMEN	1 mL Serum
TEMPERATURE	2 – 8°C
SETUP SCHEDULE	DAILY
TAT	1 day after setup schedule.
METHODOLOGY	Immunoturbidimetric
PATIENT PREPARATION	
COMMENT/S	

ASPARTATE AMINOTRANSFERASE (AST)*

CODE	1102
SPECIMEN	1 mL Serum
TEMPERATURE	2 – 8°C
SETUP SCHEDULE	DAILY
TAT	1 day after setup schedule.
METHODOLOGY	IFCC
PATIENT PREPARATION	Not Applicable
COMMENT/S	

BETA HUMAN CHORIONIC GONADOTROPHIN (βHCG)*

CODE	13072
SPECIMEN	1 mL Serum
TEMPERATURE	2 – 8°C
SETUP SCHEDULE	DAILY
TAT	1 day after setup schedule.
METHODOLOGY	Chemiluminescence Immunoassay
PATIENT PREPARATION	Not Applicable
COMMENT/S	

BICARBONATE*

CODE	1115
SPECIMEN	1 mL Serum
TEMPERATURE	2 – 8°C
SETUP SCHEDULE	DAILY
TAT	1 day after setup schedule.
METHODOLOGY	ISE, Indirect

PATIENT PREPARATION Not Applicable

COMMENT/S

BILIRUBIN (DIRECT)*

CODE	1118
SPECIMEN	1 mL Serum
TEMPERATURE	2 – 8°C
SETUP SCHEDULE	DAILY
TAT	1 day after setup schedule.
METHODOLOGY	Diazo SALT w/BLK
PATIENT PREPARATION	Not Applicable
COMMENT/S	Sample must be protected from light. Avoid hemolyzed and lipemic samples.

BILIRUBIN (INDIRECT)*

CODE	1119
SPECIMEN	1 mL Serum
TEMPERATURE	2 – 8°C
SETUP SCHEDULE	DAILY
TAT	1 day after setup schedule.
METHODOLOGY	Calculated
PATIENT PREPARATION	Not Applicable
COMMENT/S	Sample must be protected from light. Avoid hemolyzed and lipemic samples.

BILIRUBIN (TOTAL)*

CODE	1120
SPECIMEN	1 mL Serum
TEMPERATURE	2 – 8°C
SETUP SCHEDULE	DAILY
TAT	1 day after setup schedule.
METHODOLOGY	DPD
PATIENT PREPARATION	Not Applicable
COMMENT/S	Sample must be protected from light. Avoid hemolyzed and lipemic samples.

Blood Culture

CODE	1132
SPECIMEN	10 mL Blood (Adult) /1-3 mL Blood (Pediatric) in each aerobic and anaerobic bottle
TEMPERATURE	4 hours at room temperature. Must be incubated at 37°C upon receipt in laboratory.
SETUP SCHEDULE	DAILY
TAT	7 days after setup schedule.
METHODOLOGY	Aerobic, disk-diffusion
PATIENT PREPARATION	Disinfect venipuncture site. Draw blood at time of febrile episode.
COMMENT/S	

Blood Film Consultation*

CODE	10055
SPECIMEN	2 mL EDTA Whole Blood
TEMPERATURE	2 – 8°C
SETUP SCHEDULE	DAILY
TAT	1 day after setup schedule.
METHODOLOGY	
PATIENT PREPARATION	Not Applicable
COMMENT/S	

Blood Group and Rh*

CODE	1004
SPECIMEN	2 mL EDTA Whole Blood
TEMPERATURE	2 – 8°C
SETUP SCHEDULE	DAILY
TAT	1 day after setup schedule.
METHODOLOGY	Haemagglutination
PATIENT PREPARATION	Not Applicable
COMMENT/S	

BLOOD UREA NITROGEN (BUN)*

CODE	1132
SPECIMEN	1 mL Serum
TEMPERATURE	2 – 8°C
SETUP SCHEDULE	DAILY
TAT	1 day after setup schedule.
METHODOLOGY	Calculated
PATIENT PREPARATION	
COMMENT/S	

C – REACTIVE PROTEIN (CRP) QUANTITATIVE*

CODE	1141
SPECIMEN	1 mL Serum
TEMPERATURE	2 – 8°C
SETUP SCHEDULE	DAILY
TAT	1 day after setup schedule.
METHODOLOGY	Immunoturbidimetric
PATIENT PREPARATION	Not Applicable
COMMENT/S	

CA 125*

CODE	1153
SPECIMEN	1 mL Serum



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TEMPERATURE 2 – 8°C
SETUP SCHEDULE AS NEEDED
TAT 1 day after setup schedule.
METHODOLOGY Chemiluminescence Immunoassay
PATIENT PREPARATION Not Applicable
COMMENT/S

CA15 – 3*

CODE 1154
SPECIMEN 1 mL Serum
TEMPERATURE 2 – 8°C
SETUP SCHEDULE AS NEEDED
TAT 1 day after setup schedule.
METHODOLOGY Chemiluminescence Immunoassay
PATIENT PREPARATION Not Applicable
COMMENT/S

CA19 – 9*

CODE 1155
SPECIMEN 1 mL Serum
TEMPERATURE 2 – 8°C
SETUP SCHEDULE AS NEEDED
TAT 1 day after setup schedule.
METHODOLOGY Chemiluminescence Immunoassay
PATIENT PREPARATION Not Applicable
COMMENT/S

CALCIUM IN SERUM (IONIZED)*

CODE 1159
SPECIMEN 1 mL Serum
TEMPERATURE 2 – 8°C
SETUP SCHEDULE DAILY
TAT 1 day after setup schedule.
METHODOLOGY Calculated
PATIENT PREPARATION
COMMENT/S

CALCIUM, TOTAL*

CODE 1160
SPECIMEN 1 mL Serum
TEMPERATURE 2 – 8°C
SETUP SCHEDULE DAILY
TAT 1 day after setup schedule.
METHODOLOGY Cresolphthalein Method

PATIENT PREPARATION Not Applicable
COMMENT/S

CARCINOEMBRYONIC ANTIGEN (CEA)*

CODE 1166
SPECIMEN 1 mL Serum
TEMPERATURE 2 – 8°C
SETUP SCHEDULE AS NEEDED
TAT 1 day after setup schedule.
METHODOLOGY Chemiluminescence Immunoassay
PATIENT PREPARATION Not Applicable
COMMENT/S

CHLORIDE IN SERUM*

CODE 1177
SPECIMEN 1 mL Serum
TEMPERATURE 2 – 8°C
SETUP SCHEDULE DAILY
TAT 1 day after setup schedule.
METHODOLOGY ISE, Indirect
PATIENT PREPARATION Not Applicable
COMMENT/S

CHOLESTEROL, TOTAL*

CODE 5252
SPECIMEN 1 mL Serum
TEMPERATURE 2 – 8°C
SETUP SCHEDULE DAILY
TAT 1 day after setup schedule.
METHODOLOGY Enzymatic Colorimetric
PATIENT PREPARATION
COMMENT/S

CMV IgG

CODE 1188
SPECIMEN 1 mL Serum
TEMPERATURE 2 – 8°C
SETUP SCHEDULE AS NEEDED
TAT 1 day after setup schedule.
METHODOLOGY Chemiluminescence Immunoassay
PATIENT PREPARATION Not Applicable
COMMENT/S

CMV IgM



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CODE 1189
SPECIMEN 1 mL Serum
TEMPERATURE 2 – 8°C
SETUP SCHEDULE AS NEEDED
TAT 1 day after setup schedule.
METHODOLOGY Chemiluminescence Immunoassay
PATIENT PREPARATION Not Applicable
COMMENT/S

Complete Blood Count (CBC)*

CODE 1190
SPECIMEN 2 mL EDTA Whole Blood
TEMPERATURE 2 – 8°C
SETUP SCHEDULE DAILY
TAT 1 day after setup schedule.
METHODOLOGY
PATIENT PREPARATION Not Applicable
COMMENT/S

CORTISOL IN SERUM (AM)*

CODE 1197
SPECIMEN 1 mL Serum
TEMPERATURE 2 – 8°C
SETUP SCHEDULE AS NEEDED
TAT 1 day after setup schedule.
METHODOLOGY Chemiluminescence Immunoassay
PATIENT PREPARATION Not Applicable
COMMENT/S Sample should be collected before 9AM.

CORTISOL IN SERUM (PM)*

CODE 1198
SPECIMEN 1 mL Serum
TEMPERATURE 2 – 8°C
SETUP SCHEDULE AS NEEDED
TAT 1 day after setup schedule.
METHODOLOGY Chemiluminescence Immunoassay
PATIENT PREPARATION Not Applicable
COMMENT/S Sample should be collected after 9PM.

COVID - IgG

CODE 14289
SPECIMEN 1 mL Serum
TEMPERATURE 2 – 8°C
SETUP SCHEDULE DAILY

TAT 1 day after setup schedule.
METHODOLOGY Chemiluminescence Immunoassay
PATIENT PREPARATION Not Applicable
COMMENT/S

COVID - IgM

CODE 14292
SPECIMEN 1 mL Serum
TEMPERATURE 2 – 8°C
SETUP SCHEDULE DAILY
TAT 1 day after setup schedule.
METHODOLOGY Chemiluminescence Immunoassay
PATIENT PREPARATION Not Applicable
COMMENT/S

CREATININE CLEARANCE

CODE
SPECIMEN 1 mL Serum + 5 mL of 24 hour urine sample
TEMPERATURE 2 – 8°C
SETUP SCHEDULE DAILY
TAT 1 day after setup schedule.
METHODOLOGY Enzymatic Colorimetric
PATIENT PREPARATION At early morning discard the first sample of urine and then record the time on the container. Collect all urine specimens in a bottle for the next 24 hours until the next morning at the same time of the first collected sample the day before. Keep the container refrigerated during the whole process.
COMMENT/S Total urine volume is required.

CREATININE IN SERUM*

CODE 1205
SPECIMEN 1 mL Serum
TEMPERATURE 2 – 8°C
SETUP SCHEDULE DAILY
TAT 1 day after setup schedule.
METHODOLOGY Enzymatic Colorimetric
PATIENT PREPARATION Not Applicable
COMMENT/S

Creatine Kinase (CK) Total*

CODE 1202
SPECIMEN 1 mL Serum
TEMPERATURE 2 – 8°C
SETUP SCHEDULE DAILY
TAT 1 day after setup schedule.
METHODOLOGY IFCC



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PATIENT PREPARATION Not Applicable

COMMENT/S

CREATINE KINASE MB (CK – MB)*

CODE 1203
SPECIMEN 1 mL Serum
TEMPERATURE 2 – 8°C
SETUP SCHEDULE AS NEEDED
TAT 1 day after setup schedule.
METHODOLOGY IFCC
PATIENT PREPARATION Not Applicable
COMMENT/S

CREATININE IN URINE*

CODE 5279
SPECIMEN 5 mL of first morning urine
TEMPERATURE 2 – 8°C
SETUP SCHEDULE DAILY
TAT 1 day after setup schedule.
METHODOLOGY Enzymatic Colorimetric
PATIENT PREPARATION Not Applicable
COMMENT/S

Culture & Examination – Ear 1st Swab*

CODE 215
SPECIMEN 1 mL fluid or swab in transport media
TEMPERATURE Ambient
SETUP SCHEDULE DAILY
TAT 5 days after setup schedule.
METHODOLOGY Aerobic, disk-diffusion
PATIENT PREPARATION Not Applicable
COMMENT/S

Culture & Examination – Ear Right*

CODE 214
SPECIMEN 1 mL fluid or swab in transport media
TEMPERATURE Ambient
SETUP SCHEDULE DAILY
TAT 5 days after setup schedule.
METHODOLOGY Aerobic, disk-diffusion
PATIENT PREPARATION Not Applicable
COMMENT/S

Culture & Examination – Eye Left*

CODE 179
SPECIMEN One swab in transport media. Two swab if gram stain is requested.
TEMPERATURE Ambient
SETUP SCHEDULE DAILY
TAT 5 days after setup schedule.
METHODOLOGY Aerobic, disk-diffusion
PATIENT PREPARATION Not Applicable
COMMENT/S

Culture & Examination – HVS*

CODE
SPECIMEN Discharge aspirate or swab, at least two swabs are needed. Use speculum without lubricant.
TEMPERATURE Ambient, transport media for one two swabs
SETUP SCHEDULE DAILY
TAT 5 days after setup schedule.
METHODOLOGY Aerobic, disk-diffusion
PATIENT PREPARATION Not Applicable
COMMENT/S No antibiotics for 48 hrs. No sexual intercourse for 2 days. Don't wash the vagina before the examination. Sample collected by treating physician.

Culture & Examination – Nasal Swab*

CODE 203
SPECIMEN Swab in transport media
TEMPERATURE Ambient
SETUP SCHEDULE DAILY
TAT 5 days after setup schedule.
METHODOLOGY Aerobic, disk-diffusion
PATIENT PREPARATION Not Applicable
COMMENT/S

Culture & Examination – Sputum*

CODE 196
SPECIMEN Expecterated sputum sample free from saliva
TEMPERATURE 2 – 8°C
SETUP SCHEDULE DAILY
TAT 5 days after setup schedule.
METHODOLOGY Aerobic, disk-diffusion
PATIENT PREPARATION Not Applicable
COMMENT/S

Culture & Examination – Stool*

CODE 195
SPECIMEN Fresh stool sample in a sterile container at 2 – 8°C or Swab in transport media at room temperature.



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TEMPERATURE	2 – 8°C
SETUP SCHEDULE	DAILY
TAT	5 days after setup schedule.
METHODOLOGY	Aerobic, disk-diffusion
PATIENT PREPARATION	No antibiotics for 48 hours. Send the sample within 2 hours. Samples in diaper are not accepted.
COMMENT/S	

Culture & Examination – Urethral Swab*

CODE	
SPECIMEN	Discharge in sterile container or air dried smear
TEMPERATURE	2 – 8°C
SETUP SCHEDULE	DAILY
TAT	5 days after setup schedule.
METHODOLOGY	Aerobic, disk-diffusion
PATIENT PREPARATION	Not Applicable
COMMENT/S	No antibiotics for 48 hours. No release of urine before the examination. Sample collected by treating physician.

Culture & Examination – Urine*

CODE	189
SPECIMEN	Clean voided midstream urine
TEMPERATURE	2 – 8°C
SETUP SCHEDULE	DAILY
TAT	3 days after setup schedule.
METHODOLOGY	Aerobic, disk-diffusion
PATIENT PREPARATION	Not Applicable
COMMENT/S	No antibiotics for 48 hours. Wash genitals with soap and water, and then collect urine (midstream) in a sterile cup. If done at home, send the sample to the lab within 2 hours.

Culture & Examination – Wound*

CODE	188
SPECIMEN	Pus or secretion in a sterile container, or two swabs at 2 – 8°C or transport media at room temperature. Pus submitted to the laboratory in a syringe for anaerobic culture must be well closed without air.
TEMPERATURE	2 – 8°C
SETUP SCHEDULE	DAILY
TAT	5 days after setup schedule.
METHODOLOGY	Aerobic, disk-diffusion
PATIENT PREPARATION	Not Applicable
COMMENT/S	Indicate site/source.

DHEA – S*

CODE	1222
SPECIMEN	1 mL Serum
TEMPERATURE	2 – 8°C

SETUP SCHEDULE	AS NEEDED
TAT	1 day after setup schedule.
METHODOLOGY	Chemiluminescence Immunoassay
PATIENT PREPARATION	Not Applicable
COMMENT/S	

Erythrocyte Sedimentation Rate (ESR)*

CODE	1236
SPECIMEN	2 mL Citrated Plasma (1:4)
TEMPERATURE	2 – 8°C
SETUP SCHEDULE	DAILY
TAT	1 day after setup schedule.
METHODOLOGY	Modified Westergren
PATIENT PREPARATION	
COMMENT/S	

ESTIMATED GLOMERULAR FILTRATION RATE (eGFR)

CODE	1236
SPECIMEN	1 mL Serum
TEMPERATURE	2 – 8°C
SETUP SCHEDULE	DAILY
TAT	1 day after setup schedule.
METHODOLOGY	Calculated
PATIENT PREPARATION	
COMMENT/S	

ESTRADIOL (E2)*

CODE	1238
SPECIMEN	1 mL Serum
TEMPERATURE	2 – 8°C
SETUP SCHEDULE	DAILY
TAT	1 day after setup schedule.
METHODOLOGY	Chemiluminescence Immunoassay
PATIENT PREPARATION	Not Applicable
COMMENT/S	

FERRITIN IN SERUM*

CODE	1256
SPECIMEN	1 mL Serum
TEMPERATURE	2 – 8°C
SETUP SCHEDULE	DAILY
TAT	1 day after setup schedule.
METHODOLOGY	Chemiluminescence Immunoassay
PATIENT PREPARATION	Not Applicable



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COMMENT/S

FOLIC ACID IN SERUM*

CODE	1261
SPECIMEN	1 mL Serum
TEMPERATURE	2 – 8°C
SETUP SCHEDULE	DAILY
TAT	1 day after setup schedule.
METHODOLOGY	Chemiluminescence Immunoassay
PATIENT PREPARATION	Not Applicable

COMMENT/S

FOLLICLE STIMULATING HORMONE (FSH)*

CODE	1262
SPECIMEN	1 mL Serum
TEMPERATURE	2 – 8°C
SETUP SCHEDULE	DAILY
TAT	1 day after setup schedule.
METHODOLOGY	Chemiluminescence Immunoassay
PATIENT PREPARATION	Not Applicable

COMMENT/S

FOOD INTOLERANCE (FOOD PRINT)

CODE	1263
SPECIMEN	3 mL Serum
TEMPERATURE	2 – 8°C
SETUP SCHEDULE	THURS
TAT	1 day after setup schedule.
METHODOLOGY	
PATIENT PREPARATION	Not Applicable

COMMENT/S

FOOD PANEL – ALLERGEN PROFILE 20

CODE	1029
SPECIMEN	3 mL Serum
TEMPERATURE	2 – 8°C
SETUP SCHEDULE	THURS
TAT	1 day after setup schedule.
METHODOLOGY	
PATIENT PREPARATION	Not Applicable

COMMENT/S

FUNGAL EXAMINATION – HAIR*

CODE	1272
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SPECIMEN	Hair trimmings
TEMPERATURE	Ambient
SETUP SCHEDULE	DAILY
TAT	1 day after setup schedule.
METHODOLOGY	KOH wet Mount
PATIENT PREPARATION	Not Applicable

COMMENT/S

FUNGAL EXAMINATION – SKIN*

CODE	1273
SPECIMEN	Skin Scrappings
TEMPERATURE	Ambient
SETUP SCHEDULE	DAILY
TAT	1 day after setup schedule.
METHODOLOGY	KOH wet Mount
PATIENT PREPARATION	Not Applicable

COMMENT/S

FUNGAL EXAMINATION – SCALP*

CODE	
SPECIMEN	Scalp Scrappings
TEMPERATURE	Ambient
SETUP SCHEDULE	DAILY
TAT	1 day after setup schedule.
METHODOLOGY	KOH wet Mount
PATIENT PREPARATION	Not Applicable

COMMENT/S

FUNGAL EXAMINATION – NAIL*

CODE	
SPECIMEN	Nail clippings
TEMPERATURE	Ambient
SETUP SCHEDULE	DAILY
TAT	1 day after setup schedule.
METHODOLOGY	KOH wet Mount
PATIENT PREPARATION	Not Applicable

COMMENT/S

G6PD Activity (Quantitative)

CODE	1275
SPECIMEN	2 mL EDTA Whole Blood
TEMPERATURE	2 – 8°C
SETUP SCHEDULE	AS NEEDED
TAT	1 day after setup schedule.



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METHODOLOGY Colorimetric
PATIENT PREPARATION Not Applicable
COMMENT/S

GAMMA GLUTAMYL TRANSFERASE (GGT)*

CODE 1277
SPECIMEN 1 mL Serum
TEMPERATURE 2 – 8°C
SETUP SCHEDULE DAILY
TAT 1 day after setup schedule.
METHODOLOGY IFCC
PATIENT PREPARATION Not Applicable
COMMENT/S

GLOBULIN IN SERUM

CODE 1283
SPECIMEN 1 mL Serum
TEMPERATURE 2 – 8°C
SETUP SCHEDULE DAILY
TAT 1 day after setup schedule.
METHODOLOGY Calculated
PATIENT PREPARATION Not Applicable
COMMENT/S

GLUCOSE IN PLASMA (2 HOUR POST PRANDIAL)*

CODE 1286
SPECIMEN 1 mL Fluoride Plasma
TEMPERATURE 2 – 8°C
SETUP SCHEDULE DAILY
TAT 1 day after setup schedule.
METHODOLOGY Enzymatic (Hexokinase)
PATIENT PREPARATION
COMMENT/S

GLUCOSE IN PLASMA (FASTING)*

CODE 1285
SPECIMEN 1 mL Serum
TEMPERATURE 2 – 8°C
SETUP SCHEDULE DAILY
TAT 1 day after setup schedule.
METHODOLOGY Enzymatic (Hexokinase)
PATIENT PREPARATION Fasting for 6 to 8 hours.
COMMENT/S

GLUCOSE IN PLASMA (RANDOM)*

CODE 1287
SPECIMEN 1 mL Serum
TEMPERATURE 2 – 8°C
SETUP SCHEDULE DAILY
TAT 1 day after setup schedule.
METHODOLOGY Enzymatic (Hexokinase)
PATIENT PREPARATION Not Applicable
COMMENT/S

GLUCOSE TOLERANCE TEST – 100g OGTT (PREGNANT)*

CODE 1288
SPECIMEN 4 x 1 mL Fluoride Plasma + 4 x 5 mL Urine
TEMPERATURE 2 – 8°C
SETUP SCHEDULE DAILY
TAT 1 day after setup schedule.
METHODOLOGY
PATIENT PREPARATION Please contact the laboratory.
COMMENT/S

GLUCOSE TOLERANCE TEST – 50g OGTT (PREGNANT)*

CODE 5239
SPECIMEN 1 mL Fluoride Plasma
TEMPERATURE 2 – 8°C
SETUP SCHEDULE DAILY
TAT 1 day after setup schedule.
METHODOLOGY Enzymatic (Hexokinase)
PATIENT PREPARATION Sample should be collected after exactly 1 hour from intake of 50 gram glucose. Patient should not be fasting.
COMMENT/S

GLUCOSE TOLERANCE TEST – 75g OGTT (NON PREGNANT)*

CODE 1463
SPECIMEN 3 x 1 mL Fluoride Plasma + 3 x 5 mL Urine
TEMPERATURE 2 – 8°C
SETUP SCHEDULE DAILY
TAT 1 day after setup schedule.
METHODOLOGY Enzymatic (Hexokinase)
PATIENT PREPARATION
COMMENT/S

GLUCOSE TOLERANCE TEST – 75g OGTT (PREGNANT)*

CODE 1290
SPECIMEN 3 x 1 mL Fluoride Plasma + 3 x 5 mL Urine
TEMPERATURE 2 – 8°C



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SETUP SCHEDULE DAILY
TAT 1 day after setup schedule.
METHODOLOGY Enzymatic (Hexokinase)

PATIENT PREPARATION

COMMENT/S

GLYCATED HAEMOGLOBIN (HbA1c)*

CODE 1291
SPECIMEN 2 mL EDTA Whole Blood
TEMPERATURE 2 – 8°C
SETUP SCHEDULE DAILY
TAT 1 day after setup schedule.
METHODOLOGY Turbidimetric

PATIENT PREPARATION

COMMENT/S

Hemoglobin

CODE 10060
SPECIMEN 2 mL EDTA Whole Blood
TEMPERATURE 2 – 8°C
SETUP SCHEDULE DAILY
TAT 1 day after setup schedule.
METHODOLOGY Photometric Measurement

PATIENT PREPARATION Not Applicable

COMMENT/S

Hemoglobin Electrophoresis*

CODE 1303
SPECIMEN 2 mL EDTA Whole Blood
TEMPERATURE 2 – 8°C
SETUP SCHEDULE THURS
TAT 1 day after setup schedule.
METHODOLOGY HPLC

PATIENT PREPARATION Not Applicable

COMMENT/S

Helicobacter pylori Urea Breath Test (UBT)

CODE
SPECIMEN Expanded air in UBT card
TEMPERATURE Ambient
SETUP SCHEDULE DAILY
TAT 1 day after setup schedule.
METHODOLOGY
PATIENT PREPARATION Fasting for 4 to 6 hours. No stomach antibiotic for at least 3- 7 days.

COMMENT/S Instruct patient to expire into UBT card 30 minutes after taking the labeled urea capsule. Prohibited for pregnant women.

Helicobacter pylori Abs*

CODE
SPECIMEN 1 mL Serum
TEMPERATURE 2 – 8°C
SETUP SCHEDULE DAILY
TAT 1 day after setup schedule.
METHODOLOGY Immunochromatographic
PATIENT PREPARATION Not Applicable
COMMENT/S

Helicobacter pylori Stool Antigen (HpSag)*

CODE 1301
SPECIMEN Fresh stool sample in a clean, dry container
TEMPERATURE 2 – 8°C
SETUP SCHEDULE DAILY
TAT 1 day after setup schedule.
METHODOLOGY Immunochromatographic
PATIENT PREPARATION Not Applicable
COMMENT/S

Hepatitis A Virus (HAV) IgM*

CODE 1304
SPECIMEN 1 mL Serum
TEMPERATURE 2 – 8°C
SETUP SCHEDULE AS NEEDED
TAT 1 day after setup schedule.
METHODOLOGY CHEMILUMINESCENCE IMMUNOASSAY
PATIENT PREPARATION Not Applicable
COMMENT/S

Hepatitis A Virus (HAV) Total

CODE 1305
SPECIMEN 1 mL Serum
TEMPERATURE 2 – 8°C
SETUP SCHEDULE AS NEEDED
TAT 1 day after setup schedule.
METHODOLOGY CHEMILUMINESCENCE IMMUNOASSAY
PATIENT PREPARATION Not Applicable
COMMENT/S

Hepatitis B Core (HBc) IgM



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CODE 1306
SPECIMEN 1 mL Serum
TEMPERATURE 2 – 8°C
SETUP SCHEDULE AS NEEDED
TAT 1 day after setup schedule.
METHODOLOGY CHEMILUMINESCENCE IMMUNOASSAY
PATIENT PREPARATION Not Applicable
COMMENT/S

Hepatitis B Core (HBc) Total

CODE 1307
SPECIMEN 1 mL Serum
TEMPERATURE 2 – 8°C
SETUP SCHEDULE AS NEEDED
TAT 1 day after setup schedule.
METHODOLOGY CHEMILUMINESCENCE IMMUNOASSAY
PATIENT PREPARATION Not Applicable
COMMENT/S

Hepatitis B Surface (HBs) Abs

CODE 1310
SPECIMEN 1 mL Serum
TEMPERATURE 2 – 8°C
SETUP SCHEDULE AS NEEDED
TAT 1 day after setup schedule.
METHODOLOGY CHEMILUMINESCENCE IMMUNOASSAY
PATIENT PREPARATION Not Applicable
COMMENT/S

HEPATITIS B SURFACE (HBs) Ag*

CODE 1311
SPECIMEN 1 mL Serum
TEMPERATURE 2 – 8°C
SETUP SCHEDULE DAILY
TAT 1 day after setup schedule.
METHODOLOGY Chemiluminescence Immunoassay
PATIENT PREPARATION Not Applicable
COMMENT/S

HEPATITIS C VIRUS (HCV) Abs*

CODE 1315
SPECIMEN 1 mL Serum
TEMPERATURE 2 – 8°C
SETUP SCHEDULE DAILY

TAT 1 day after setup schedule.
METHODOLOGY Chemiluminescence Immunoassay
PATIENT PREPARATION Not Applicable
COMMENT/S

HIGH DENSITY LIPOPROTEIN (HDL)*

CODE 5251
SPECIMEN 1 mL Serum
TEMPERATURE 2 – 8°C
SETUP SCHEDULE DAILY
TAT 1 day after setup schedule.
METHODOLOGY Enzymatic Colorimetric
PATIENT PREPARATION Fasting 12 – 14 hours.
COMMENT/S

HIV P24 Ag / Abs 1 & 2*

CODE 1330
SPECIMEN 1 mL Serum
TEMPERATURE 2 – 8°C
SETUP SCHEDULE DAILY
TAT 1 day after setup schedule.
METHODOLOGY Chemiluminescence Immunoassay
PATIENT PREPARATION Not Applicable
COMMENT/S

HOMA ESTIMATED INSULIN RESISTANCE

CODE 5277
SPECIMEN 1 mL Serum + 1 mL Fluoride Plasma
TEMPERATURE 2 – 8°C
SETUP SCHEDULE DAILY
TAT 1 day after setup schedule.
METHODOLOGY Calculated
PATIENT PREPARATION Not Applicable
COMMENT/S

INHALATION PANEL – ALLERGEN 20

CODE 1030
SPECIMEN 3 mL Serum
TEMPERATURE 2 – 8°C
SETUP SCHEDULE THURS
TAT 1 day after setup schedule.
METHODOLOGY
PATIENT PREPARATION Not Applicable
COMMENT/S



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Immunoglobulin E (IgE) Total*

CODE	1348
SPECIMEN	1 mL Serum
TEMPERATURE	2 – 8°C
SETUP SCHEDULE	DAILY
TAT	1 day after setup schedule.
METHODOLOGY	Immunoturbidimetric
PATIENT PREPARATION	Not Applicable
COMMENT/S	

INFLUENZA A AND B VIRUS Ag*

CODE	1388
SPECIMEN	1 mL Serum
TEMPERATURE	2 – 8°C
SETUP SCHEDULE	DAILY
TAT	1 day after setup schedule.
METHODOLOGY	Immunochromatographic
PATIENT PREPARATION	Not Applicable
COMMENT/S	

Insulin Level*

CODE	1471
SPECIMEN	1 mL Serum
TEMPERATURE	2 – 8°C
SETUP SCHEDULE	AS NEEDED
TAT	1 day after setup schedule.
METHODOLOGY	Chemiluminescence Immunoassay
PATIENT PREPARATION	Not Applicable
COMMENT/S	

Insulin (2hrs Post Prandial)*

CODE	
SPECIMEN	1 mL Serum
TEMPERATURE	2 – 8°C
SETUP SCHEDULE	AS NEEDED
TAT	1 day after setup schedule.
METHODOLOGY	Chemiluminescence Immunoassay
PATIENT PREPARATION	Not Applicable
COMMENT/S	

IRON*

CODE	1399
SPECIMEN	1 mL Serum

TEMPERATURE	2 – 8°C
SETUP SCHEDULE	DAILY
TAT	1 day after setup schedule.
METHODOLOGY	TPTZ
PATIENT PREPARATION	Fasting for 6 to 8 hours. Iron-containing supplements should be avoided for 24 hours prior to draw.
COMMENT/S	

LACTATE DEHYDROGENASE (LDH)*

CODE	1403
SPECIMEN	1 mL Serum
TEMPERATURE	2 – 8°C
SETUP SCHEDULE	DAILY
TAT	1 day after setup schedule.
METHODOLOGY	IFCC
PATIENT PREPARATION	Not Applicable
COMMENT/S	

LIPASE*

CODE	1418
SPECIMEN	1 mL Serum
TEMPERATURE	2 – 8°C
SETUP SCHEDULE	DAILY
TAT	1 day after setup schedule.
METHODOLOGY	Enzymatic Colorimetric
PATIENT PREPARATION	Not Applicable
COMMENT/S	

LIPID PROFILE

CODE	5235
SPECIMEN	1 mL Serum
TEMPERATURE	2 – 8°C
SETUP SCHEDULE	DAILY
TAT	1 day after setup schedule.
METHODOLOGY	
PATIENT PREPARATION	Fasting for 12 hours.
COMMENT/S	

LOW DENSITY LIPOPROTEIN (LDL)*

CODE	5249
SPECIMEN	1 mL Serum
TEMPERATURE	2 – 8°C
SETUP SCHEDULE	DAILY
TAT	1 day after setup schedule.
METHODOLOGY	Enzymatic Selective Protection



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PATIENT PREPARATION Not Applicable

COMMENT/S

LUTEINIZING HORMONE (LH)*

CODE 1425
SPECIMEN 1 mL Serum
TEMPERATURE 2 – 8°C
SETUP SCHEDULE DAILY
TAT 1 day after setup schedule.
METHODOLOGY Chemiluminescence Immunoassay
PATIENT PREPARATION Not Applicable
COMMENT/S

MAGNESIUM*

CODE 1428
SPECIMEN 1 mL Serum
TEMPERATURE 2 – 8°C
SETUP SCHEDULE DAILY
TAT 1 day after setup schedule.
METHODOLOGY Colorimetric
PATIENT PREPARATION Not Applicable
COMMENT/S

MALARIA Ag*

CODE 1429
SPECIMEN 1 mL Serum
TEMPERATURE 2 – 8°C
SETUP SCHEDULE DAILY
TAT 1 day after setup schedule.
METHODOLOGY Immunochromatographic
PATIENT PREPARATION Not Applicable
COMMENT/S

Malaria Blood Film*

CODE 1430
SPECIMEN 2 mL EDTA Whole Blood
TEMPERATURE 2 – 8°C
SETUP SCHEDULE DAILY
TAT 1 day after setup schedule.
METHODOLOGY
PATIENT PREPARATION Not Applicable
COMMENT/S

MICROALBUMINURIA (ALBUMIN / CREATININE RATIO)*

CODE 1438
SPECIMEN 5 mL of first morning urine
TEMPERATURE 2 – 8°C
SETUP SCHEDULE DAILY
TAT 1 day after setup schedule.
METHODOLOGY Calculated
PATIENT PREPARATION Not Applicable
COMMENT/S

Parathyroid Hormone (PTH)*

CODE 1471
SPECIMEN 1 mL Serum
TEMPERATURE 2 – 8°C
SETUP SCHEDULE AS NEEDED
TAT 1 day after setup schedule.
METHODOLOGY Chemiluminescence Immunoassay
PATIENT PREPARATION Not Applicable
COMMENT/S

Partial Thromboplastin Time (PTT)*

CODE 1472
SPECIMEN 1 mL Platelet Poor Citrated Plasma (1:9)
TEMPERATURE Frozen
SETUP SCHEDULE DAILY
TAT 1 day after setup schedule.
METHODOLOGY Clot retraction
PATIENT PREPARATION Not Applicable
COMMENT/S

PHOSPHORUS*

CODE 1480
SPECIMEN 1 mL Serum
TEMPERATURE 2 – 8°C
SETUP SCHEDULE DAILY
TAT 1 day after setup schedule.
METHODOLOGY Colorimetric
PATIENT PREPARATION Not Applicable
COMMENT/S

Post Vaccination Covid-19 Antibody Profile

CODE
SPECIMEN 1 mL Serum
TEMPERATURE 2 – 8°C
SETUP SCHEDULE SUN, TUES, WED



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TAT 1 day after setup schedule.

METHODOLOGY ELISA

PATIENT PREPARATION Not Applicable

COMMENT/S

POTASSIUM*

CODE 5224

SPECIMEN 1 mL Serum

TEMPERATURE 2 – 8°C

SETUP SCHEDULE DAILY

TAT 1 day after setup schedule.

METHODOLOGY Selective Electrode

PATIENT PREPARATION Not Applicable

COMMENT/S Avoid hemolyzed samples.

PROGESTERONE*

CODE 1490

SPECIMEN 1 mL Serum

TEMPERATURE 2 – 8°C

SETUP SCHEDULE DAILY

TAT 1 day after setup schedule.

METHODOLOGY Chemiluminescence Immunoassay

PATIENT PREPARATION Not Applicable

COMMENT/S

PROLACTIN*

CODE 1492

SPECIMEN 1 mL Serum

TEMPERATURE 2 – 8°C

SETUP SCHEDULE DAILY

TAT 1 day after setup schedule.

METHODOLOGY Chemiluminescence Immunoassay

PATIENT PREPARATION Not Applicable

COMMENT/S

PROSTATE SPECIFIC ANTIGEN (PSA), FREE*

CODE

SPECIMEN 1 mL Serum

TEMPERATURE 2 – 8°C

SETUP SCHEDULE DAILY

TAT 1 day after setup schedule.

METHODOLOGY Chemiluminescence Immunoassay

PATIENT PREPARATION Not Applicable

COMMENT/S

PROSTATE SPECIFIC ANTIGEN (PSA), TOTAL AND FREE*

CODE 14113

SPECIMEN 1 mL Serum

TEMPERATURE 2 – 8°C

SETUP SCHEDULE DAILY

TAT 1 day after setup schedule.

METHODOLOGY Chemiluminescence Immunoassay

PATIENT PREPARATION Not Applicable

COMMENT/S

Prothrombin Time (PT)*

CODE 1501

SPECIMEN 1 mL Platelet Poor Citrated Plasma (1:9)

TEMPERATURE Frozen

SETUP SCHEDULE DAILY

TAT 1 day after setup schedule.

METHODOLOGY Clot retraction

PATIENT PREPARATION Not Applicable

COMMENT/S

Pus Examination C/S

CODE 484

SPECIMEN Fresh prostatic fluid sample in sterile container

TEMPERATURE

SETUP SCHEDULE: DAILY

TAT 5 days after setup schedule.

PATIENT PREP Not Applicable

COMMENTS Indicate site/source.

Rapid Plasma Reagin (RPR)*

CODE 1504

SPECIMEN 1 mL Serum

TEMPERATURE 4 °C for 24 hours / -20°C for 1 month

SETUP SCHEDULE DAILY

TAT 1 day after setup schedule.

METHODOLOGY Flocculation card test

PATIENT PREPARATION Not Applicable

COMMENT/S

RESPIRATORY SYNCYTIAL VIRUS (RSV)*

CODE

SPECIMEN Throat swab

TEMPERATURE Ambient

SETUP SCHEDULE DAILY

TAT 1 day after setup schedule.



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METHODOLOGY Immunochromatographic

PATIENT PREPARATION Not Applicable

COMMENT/S

VPIII - VAGINAL MICROBIAL IDENTIFICATION*

CODE 1640
SPECIMEN Female genital swab
TEMPERATURE Ambient
SETUP SCHEDULE DAILY
TAT 1 day after setup schedule.
METHODOLOGY Direct test DNA probe based detection identification
PATIENT PREPARATION Not Applicable
COMMENT/S

Reticulocyte Count*

CODE 10057
SPECIMEN 2 mL EDTA Whole Blood
TEMPERATURE 2 – 8°C
SETUP SCHEDULE DAILY
TAT 1 day after setup schedule.
METHODOLOGY
PATIENT PREPARATION Not Applicable
COMMENT/S

Rheumatoid Factor (RF) Quantitative

CODE 736

SPECIMEN 1 mL Serum
TEMPERATURE 2 – 8°C
SETUP SCHEDULE: SAT, MON, WED
TAT 1 day after setup schedule.
PATIENT PREP Not Applicable
COMMENTS

RHEUMATOID FACTOR (RF) QUANTITATIVE*

CODE 1513
SPECIMEN 1 mL Serum
TEMPERATURE 2 – 8°C
SETUP SCHEDULE AS NEEDED
TAT 1 day after setup schedule.
METHODOLOGY Immunoturbidimetric
PATIENT PREPARATION Not Applicable
COMMENT/S

Rotavirus Antigen*

CODE 1514

SPECIMEN Fresh stool sample in a clean, dry container at 2 – 8°C

TEMPERATURE Ambient

SETUP SCHEDULE DAILY

TAT 1 day after setup schedule.

METHODOLOGY Immune chromatography

PATIENT PREPARATION Not Applicable

COMMENT/S

RUBELLA IgG*

CODE 1515
SPECIMEN 1 mL Serum
TEMPERATURE 2 – 8°C
SETUP SCHEDULE AS NEEDED
TAT 1 day after setup schedule.
METHODOLOGY Chemiluminescence Immunoassay
PATIENT PREPARATION Not Applicable
COMMENT/S

RUBELLA IgM*

CODE 1516
SPECIMEN 1 mL Serum
TEMPERATURE 2 – 8°C
SETUP SCHEDULE AS NEEDED
TAT 1 day after setup schedule.
METHODOLOGY Chemiluminescence Immunoassay
PATIENT PREPARATION Not Applicable
COMMENT/S

Semen Examination

CODE 1520
SPECIMEN Fresh semen sample in sterile container
TEMPERATURE Submit to the laboratory as soon as possible
SETUP SCHEDULE: DAILY
TAT 1 day after setup schedule.
METHODOLOGY
PATIENT PREP No copulation or wet dreams for a period of 2 – 7 days.
COMMENTS Useful in the diagnosis of infertility/suspected infection.

SEX HORMONE BINDING GLOBULIN (SHBG)*

CODE 1525
SPECIMEN 1 mL Serum
TEMPERATURE 2 – 8°C
SETUP SCHEDULE DAILY
TAT 1 day after setup schedule.



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METHODOLOGY Chemiluminescence Immunoassay

PATIENT PREPARATION Not Applicable

COMMENT/S

Sickle Cell Solubility Test*

CODE 10078
SPECIMEN 2 mL EDTA Whole Blood
TEMPERATURE 2 – 8°C
SETUP SCHEDULE DAILY
TAT 1 day after setup schedule.
METHODOLOGY Modified Nalbandian
PATIENT PREPARATION Not Applicable
COMMENT/S

SODIUM IN SERUM*

CODE 1532
SPECIMEN 1 mL Serum
TEMPERATURE 2 – 8°C
SETUP SCHEDULE DAILY
TAT 1 day after setup schedule.
METHODOLOGY ISE, indirect
PATIENT PREPARATION Not Applicable
COMMENT/S

STREPTOCOCCAL A RAPID TEST (THROAT SWAB)*

CODE 1545
SPECIMEN Throat swab
TEMPERATURE Ambient
SETUP SCHEDULE DAILY
TAT 1 day after setup schedule.
METHODOLOGY Immunochromatographic
PATIENT PREPARATION Not Applicable
COMMENT/S

Occult Blood*

CODE 1462
SPECIMEN Fresh stool sample in a clean, dry container
TEMPERATURE 2 – 8°C
SETUP SCHEDULE DAILY
TAT 1 day after setup schedule.
METHODOLOGY Immunochromatographic
PATIENT PREPARATION Not Applicable
COMMENT/S

Stool Examination*

CODE 6008
SPECIMEN Fresh stool sample in a clean, dry container at 2 – 8°C
TEMPERATURE Ambient
SETUP SCHEDULE DAILY
TAT 1 day after setup schedule.
METHODOLOGY Visual, Microscopy
PATIENT PREPARATION Not Applicable
COMMENT/S Samples in diaper are not accepted.

T3, FREE*

CODE 1268
SPECIMEN 1 mL Serum
TEMPERATURE 2 – 8°C
SETUP SCHEDULE DAILY
TAT 1 day after setup schedule.
METHODOLOGY Chemiluminescence Immunoassay
PATIENT PREPARATION Not Applicable
COMMENT/S

T4, TOTAL

CODE 1552
SPECIMEN 1 mL Serum
TEMPERATURE 2 – 8°C
SETUP SCHEDULE AS NEEDED
TAT 1 day after setup schedule.
METHODOLOGY Chemiluminescence Immunoassay
PATIENT PREPARATION Not Applicable
COMMENT/S

T4, FREE*

CODE 1269
SPECIMEN 1 mL Serum
TEMPERATURE 2 – 8°C
SETUP SCHEDULE DAILY
TAT 1 day after setup schedule.
METHODOLOGY Chemiluminescence Immunoassay
PATIENT PREPARATION Not Applicable
COMMENT/S

T4, TOTAL*

CODE 1554
SPECIMEN 1 mL Serum



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TEMPERATURE 2 – 8°C
SETUP SCHEDULE AS NEEDED
TAT 1 day after setup schedule.
METHODOLOGY Chemiluminescence Immunoassay
PATIENT PREPARATION Not Applicable
COMMENT/S

TESTOSTERONE, FREE

CODE 13028
SPECIMEN 1 mL Serum
TEMPERATURE 2 – 8°C
SETUP SCHEDULE DAILY
TAT 1 day after setup schedule.
METHODOLOGY Calculated
PATIENT PREPARATION Not Applicable
COMMENT/S

TESTOSTERONE, TOTAL*

CODE 1559
SPECIMEN 1 mL Serum
TEMPERATURE 2 – 8°C
SETUP SCHEDULE DAILY
TAT 1 day after setup schedule.
METHODOLOGY Chemiluminescence Immunoassay
PATIENT PREPARATION Not Applicable
COMMENT/S

Culture & Examination – Throat*

CODE 193
SPECIMEN Two swabs are needed
TEMPERATURE Ambient, transport media for one of the two swab
SETUP SCHEDULE DAILY
TAT 5 days after setup schedule.
METHODOLOGY
PATIENT PREPARATION Not Applicable
COMMENT/S

THYROGLOBULIN

CODE 1564
SPECIMEN 1 mL Serum
TEMPERATURE 2 – 8°C
SETUP SCHEDULE AS NEEDED
TAT 1 day after setup schedule.
METHODOLOGY Chemiluminescence Immunoassay

PATIENT PREPARATION Not Applicable
COMMENT/S

THYROID STIMULATING HORMONE (TSH)*

CODE 1566
SPECIMEN 1 mL Serum
TEMPERATURE 2 – 8°C
SETUP SCHEDULE DAILY
TAT 1 day after setup schedule.
METHODOLOGY Chemiluminescence Immunoassay
PATIENT PREPARATION Not Applicable
COMMENT/S

TOTAL IRON BINDING CAPACITY (TIBC)*

CODE 1573
SPECIMEN 1 mL Serum
TEMPERATURE 2 – 8°C
SETUP SCHEDULE DAILY
TAT 1 day after setup schedule.
METHODOLOGY Calculated
PATIENT PREPARATION Not Applicable
COMMENT/S

TOTAL PROTEIN*

CODE 1574
SPECIMEN 1 mL Serum
TEMPERATURE 2 – 8°C
SETUP SCHEDULE DAILY
TAT 1 day after setup schedule.
METHODOLOGY Biuret
PATIENT PREPARATION Not Applicable
COMMENT/S

TOXOPLASMA IgG

CODE 1577
SPECIMEN 1 mL Serum
TEMPERATURE 2 – 8°C
SETUP SCHEDULE DAILY
TAT 1 day after setup schedule.
METHODOLOGY Chemiluminescence Immunoassay
PATIENT PREPARATION Not Applicable
COMMENT/S

TOXOPLASMA IgM



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CODE 1578
SPECIMEN 1 mL Serum
TEMPERATURE 2 – 8°C
SETUP SCHEDULE DAILY
TAT 1 day after setup schedule.
METHODOLOGY Chemiluminescence Immunoassay
PATIENT PREPARATION Not Applicable
COMMENT/S

TRANSFERRIN

CODE 1580
SPECIMEN 1 mL Serum
TEMPERATURE 2 – 8°C
SETUP SCHEDULE DAILY
TAT 1 day after setup schedule.
METHODOLOGY Immunoturbidimetric
PATIENT PREPARATION Not Applicable
COMMENT/S

TRIGLYCERIDES*

CODE 5250
SPECIMEN 1 mL Serum
TEMPERATURE 2 – 8°C
SETUP SCHEDULE DAILY
TAT 1 day after setup schedule.
METHODOLOGY Enzymatic Colorimetric
PATIENT PREPARATION Not Applicable
COMMENT/S Fasting 12 – 14 hours

T3, TOTAL*

CODE 1552
SPECIMEN 1 mL Serum
TEMPERATURE 2 – 8°C
SETUP SCHEDULE AS NEEDED
TAT 1 day after setup schedule.
METHODOLOGY Chemiluminescence Immunoassay
PATIENT PREPARATION Not Applicable
COMMENT/S

TROPONIN I – HIGH SENSITIVE*

CODE 1588
SPECIMEN 1 mL Serum
TEMPERATURE 2 – 8°C
SETUP SCHEDULE DAILY

TAT 1 day after setup schedule.
METHODOLOGY Chemiluminescence Immunoassay
PATIENT PREPARATION Not Applicable
COMMENT/S

UREA IN SERUM*

CODE 1595
SPECIMEN 1 mL Serum
TEMPERATURE 2 – 8°C
SETUP SCHEDULE DAILY
TAT 1 day after setup schedule.
METHODOLOGY Kinetic Urease with GLDH
PATIENT PREPARATION Not Applicable
COMMENT/S

URIC ACID IN SERUM*

CODE 1603
SPECIMEN 1 mL Serum
TEMPERATURE 2 – 8°C
SETUP SCHEDULE DAILY
TAT 1 day after setup schedule.
METHODOLOGY Trinder
PATIENT PREPARATION Not Applicable
COMMENT/S

Urine Analysis*

CODE 1605
SPECIMEN 15 mL Urine
TEMPERATURE 2 – 8°C
SETUP SCHEDULE DAILY
TAT 1 day after setup schedule.
METHODOLOGY Dipstick
PATIENT PREPARATION Not Applicable
COMMENT/S Clean catch voided urine.

VDRL

CODE 1621
SPECIMEN 1 mL Serum
TEMPERATURE 4 °C for 24 hours / -20°C for 1 month
SETUP SCHEDULE DAILY
TAT 1 day after setup schedule.
METHODOLOGY Latex Agglutination
PATIENT PREPARATION Not Applicable
COMMENT/S



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VITAMIN B12 (CYANOCOBALAMIN)*

CODE	1624
SPECIMEN	1 mL Serum
TEMPERATURE	2 – 8°C
SETUP SCHEDULE	DAILY
TAT	1 day after setup schedule.
METHODOLOGY	Chemiluminescence Immunoassay
PATIENT PREPARATION	Not Applicable
COMMENT/S	

VITAMIN D (25 HYDROXYCHOLECALCIFEROL)*

CODE	1631
SPECIMEN	1 mL Serum
TEMPERATURE	2 – 8°C
SETUP SCHEDULE	DAILY
TAT	1 day after setup schedule.
METHODOLOGY	Chemiluminescence Immunoassay
PATIENT PREPARATION	Not Applicable
COMMENT/S	

WIDAL TEST*

CODE	1644
SPECIMEN	1 mL Serum
TEMPERATURE	2 – 8°C
SETUP SCHEDULE	DAILY
TAT	1 day after setup schedule.
METHODOLOGY	Card agglutination method
PATIENT PREPARATION	Not Applicable
COMMENT/S	

ZINC

CODE	1648
SPECIMEN	1 mL Serum
TEMPERATURE	2 – 8°C
SETUP SCHEDULE	DAILY
TAT	1 day after setup schedule.
METHODOLOGY	Direct Colorimetric
PATIENT PREPARATION	Not Applicable
COMMENT/S	

HISTOPATHOLOGY AND CYTOLOGY TEST MENU

Tests marked with "*" are under ISO 15189:2012 Scope of Accreditation.

Biopsy (Large)

CODE	12012
SPECIMEN	Large size tissue biopsy fixed in 10% formalin
TEMPERATURE	Ambient
SETUP SCHEDULE	DAILY
TAT	5 days after setup schedule.
METHODOLOGY	Microscopic examination of formalin fixed, processed tissue and preparation of H&E stained slides from paraffin embedded sections
PATIENT PREPARATION	Not Applicable
COMMENT/S	Complete clinical data must be submitted with the specimen.

Biopsy (Medium)

CODE	12013
SPECIMEN	Medium size tissue biopsy fixed in 10% formalin
TEMPERATURE	Ambient
SETUP SCHEDULE	DAILY
TAT	5 days after setup schedule.
METHODOLOGY	Microscopic examination of formalin fixed, processed tissue and preparation of H&E stained slides from paraffin embedded sections
PATIENT PREPARATION	Not Applicable
COMMENT/S	Complete clinical data must be submitted with the specimen.

Biopsy (Small)

CODE	12014
SPECIMEN	Small size tissue biopsy fixed in 10% formalin (ex. Endoscopy Biopsy, D & C, Appendix)
TEMPERATURE	Ambient
SETUP SCHEDULE	DAILY
TAT	5 days after setup schedule.
METHODOLOGY	Microscopic examination of formalin fixed, processed tissue and preparation of H&E stained slides from paraffin embedded sections
PATIENT PREPARATION	Not Applicable
COMMENT/S	Complete clinical data must be submitted with the specimen. Only Testicular Biopsy needs to be fixed in Bouin's solution not formalin.

Biopsy Level III

CODE	1126
SPECIMEN	Biopsy Level IV fixed in 10% formalin
TEMPERATURE	Ambient
SETUP SCHEDULE	DAILY
TAT	6 days after setup schedule.
METHODOLOGY	Microscopic examination of formalin fixed, processed tissue and preparation of H&E stained slides from paraffin embedded sections
PATIENT PREPARATION	Not Applicable



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COMMENT/S Complete clinical data must be submitted with the specimen.

COMMENT/S Complete clinical data must be submitted with the specimen.

Biopsy Level IV

CODE	1127
SPECIMEN	Biopsy Level IV fixed in 10% formalin
TEMPERATURE	Ambient
SETUP SCHEDULE	DAILY
TAT	6 days after setup schedule.
METHODOLOGY	Microscopic examination of formalin fixed, processed tissue and preparation of H&E stained slides from paraffin embedded sections
PATIENT PREPARATION	Not Applicable
COMMENT/S	Complete clinical data must be submitted with the specimen.

Biopsy Level V

CODE	1127
SPECIMEN	Biopsy Level IV fixed in 10% formalin
TEMPERATURE	Ambient
SETUP SCHEDULE	DAILY
TAT	6 days after setup schedule.
METHODOLOGY	Microscopic examination of formalin fixed, processed tissue and preparation of H&E stained slides from paraffin embedded sections
PATIENT PREPARATION	Not Applicable
COMMENT/S	Complete clinical data must be submitted with the specimen.

Liquid based Cytology (LBC) Gyn*

CODE	8001
SPECIMEN	Cervical smear fixed in liquid base vial for Gynecological Cytology using Sure Path vial
TEMPERATURE	Ambient
SETUP SCHEDULE	DAILY
TAT	4 days after setup schedule.
METHODOLOGY	Microscopic examination of PAP stain Slides
PATIENT PREPARATION	Not Applicable
COMMENT/S	Complete clinical data must be submitted with the specimen.

Liquid based Cytology (LBC) Non Gyn*

CODE	1421
SPECIMEN	20 to 50 mL Body fluid, Effusion, Aspirations, Urine, Lavages, etc. for non-Gynecological Cytology. Must be sent immediately to the laboratory or use LBC vial if delayed.
TEMPERATURE	Ambient within 1 hour / 4°C if delay is anticipated. CSF and Urine: Degenerates within 1 hour even with refrigeration.
SETUP SCHEDULE	DAILY
TAT	4 days after setup schedule.
METHODOLOGY	Microscopic examination of PAP stain Slides
PATIENT PREPARATION	Not Applicable

MOLECULAR BIOLOGY TEST MENU

Tests marked with '*' are under ISO 15189:2012 Scope of Accreditation.

Human Papilloma Virus (HPV) detection and genotyping by PCR

CODE	16018
SPECIMEN	Sure path or Thin Prep
TEMPERATURE	2 – 8°C
SETUP SCHEDULE	DAILY
TAT	1 day after setup schedule.
METHODOLOGY	RT-PCR
PATIENT PREPARATION	Not Applicable
COMMENT/S	

Molecular Detection of COVID-19 by Real Time PCR

CODE	16027
SPECIMEN	Swab on VTM
TEMPERATURE	2 – 8°C
SETUP SCHEDULE	DAILY
TAT	1 day after setup schedule.
METHODOLOGY	RT-PCR
PATIENT PREPARATION	Not Applicable
COMMENT/S	

SEND OUT TEST MENU

1,25-(OH)2 vitamin D

CODE	
SPECIMEN	1 mL Serum
TEMPERATURE	2 – 8°C
SETUP SCHEDULE	Contact Customer Care
TAT	14 days
METHODOLOGY	CIA (Chemiluminescence immunoassay)
PATIENT PREPARATION	Not Applicable
COMMENT/S	

17-hydroxyprogesterone

CODE	
SPECIMEN	1 mL Serum
TEMPERATURE	2 – 8°C
SETUP SCHEDULE	Contact Customer Care

TAT 14 days
METHODOLOGY RIA (Radioimmunoassay)
PATIENT PREPARATION Not Applicable
COMMENT/S

ACTH (AM)

CODE
SPECIMEN 1 mL EDTA Plasma
TEMPERATURE Frozen
SETUP SCHEDULE Contact Customer Care
TAT 14 days
METHODOLOGY ECLIA (Electrochemiluminescence Immunoassay)
PATIENT PREPARATION Not Applicable
COMMENT/S The plasma samples were drawn between 7-10 a.m.
Only use pre-cooled sampling vials. After drawing the blood, put the vials immediately on ice. Use a cooled centrifuge to separate the plasma. Measure samples immediately or freeze them at -20°C
Freeze only once.

Aldosterone

CODE
SPECIMEN 1 mL Serum
TEMPERATURE Frozen
SETUP SCHEDULE Contact Customer Care
TAT 14 days
METHODOLOGY LC (Liquid chromatography) / MS-MS
PATIENT PREPARATION Not Applicable
COMMENT/S Precautions when collecting blood for serum/plasma aldosterone tests
1. Sampling in the morning is recommended
- Samples which were collected in the morning tends to show higher serum/plasma aldosterone concentrations than samples which were collected in the afternoon
2. Dietary Causes of Pseudohyperaldosteronism
- Licorice (e.g., candies, drinks, gum, and tea containing licorice, etc.), grapefruit, carbenoxolone, etc.
- In particular, avoid taking licorice for at least two weeks before the test.
3. Dietary and position influences
- Avoid high or low sodium diets, stress, and strenuous exercises
- Maintain constant position before and during blood collection (samples which were collected in upright position tends to show 2 to 3 times higher serum/plasma aldosterone concentrations than samples which were collected in supine position)
4. Drug effects
- Influence on test results when taking nonsteroidal anti-inflammatory drugs (NSAIDs: ibuprofen, etc.), diuretics, beta blockers, steroids, angiotensin converting enzyme inhibitors, oral contraceptives, etc.
5. After specimens collection, the specimens should be separated by centrifugation and stored frozen immediately after centrifugation. (-20°C or below)

Amyloid A

CODE

SPECIMEN 1 mL Serum
TEMPERATURE 2 – 8°C
SETUP SCHEDULE Contact Customer Care
TAT 14 days
METHODOLOGY TIA (Turbidimetric immunoassay)
PATIENT PREPARATION Not Applicable
COMMENT/S

Anti-centromere Ab

CODE
SPECIMEN 1 mL Serum
TEMPERATURE 2 – 8°C
SETUP SCHEDULE Contact Customer Care
TAT 14 days
METHODOLOGY FEIA (Fluorescence enzyme immunoassay)
PATIENT PREPARATION Not Applicable
COMMENT/S

Anti-gastric parietal cell Ab

CODE
SPECIMEN 1 mL Serum
TEMPERATURE 2 – 8°C
SETUP SCHEDULE Contact Customer Care
TAT 14 days
METHODOLOGY Fluorescent antibody test
PATIENT PREPARATION Not Applicable
COMMENT/S

Anti-gastric parietal cell Ab

CODE
SPECIMEN 1 mL Serum
TEMPERATURE 2 – 8°C
SETUP SCHEDULE Contact Customer Care
TAT 14 days
METHODOLOGY Fluorescent antibody test
PATIENT PREPARATION Not Applicable
COMMENT/S

Anti-histone Ab

CODE
SPECIMEN 1 mL Serum
TEMPERATURE 2 – 8°C
SETUP SCHEDULE Contact Customer Care
TAT 14 days



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SETUP SCHEDULE Contact Customer Care
TAT 14 days
METHODOLOGY FEIA (Fluorescence enzyme immunoassay)
PATIENT PREPARATION Not Applicable
COMMENT/S

Biological antioxidant potential and Reactive oxygen metabolites

CODE
SPECIMEN 2 mL Serum
TEMPERATURE 2 – 8°C
SETUP SCHEDULE Contact Customer Care
TAT 14 days
METHODOLOGY Colorimetry
PATIENT PREPARATION Not Applicable
COMMENT/S

C1 inhibitor (C1 inactivator, C1-esterase inhibitor)

CODE
SPECIMEN 1 mL Serum
TEMPERATURE 2 – 8°C
SETUP SCHEDULE Contact Customer Care
TAT 14 days
METHODOLOGY NIA (Nephelometric immunoassay)
PATIENT PREPARATION Not Applicable
COMMENT/S

Complement C4

CODE
SPECIMEN 1 mL Serum
TEMPERATURE 2 – 8°C
SETUP SCHEDULE Contact Customer Care
TAT 14 days
METHODOLOGY Turbidimetric immunoassay
PATIENT PREPARATION Not Applicable
COMMENT/S In very rare cases, gammopathy, in particular type IgM (Waldenstrom’s macroglobulinemia), may cause unreliable results.

Complement C3

CODE
SPECIMEN 1 mL Serum
TEMPERATURE 2 – 8°C
SETUP SCHEDULE Contact Customer Care
TAT 14 days
METHODOLOGY Turbidimetric immunoassay

PATIENT PREPARATION Not Applicable
COMMENT/S In very rare cases, gammopathy, in particular type IgM (Waldenstrom’s macroglobulinemia), may cause unreliable results.

DHEA-S

CODE
SPECIMEN 1 mL Serum
TEMPERATURE 2 – 8°C
SETUP SCHEDULE Contact Customer Care
TAT 14 days
METHODOLOGY ECLIA (Electrochemiluminescence Immunoassay)
PATIENT PREPARATION Not Applicable
COMMENT/S

Factor II (2)

CODE
SPECIMEN 2 mL Platelet Poor Citrated Plasma (1:9)
TEMPERATURE Frozen
SETUP SCHEDULE Contact Customer Care
TAT 14 days
METHODOLOGY Clot Based
PATIENT PREPARATION Not Applicable
COMMENT/S 1. The clotted or hemolytic specimens are not adequate.
 2. Only frozen poor-platelet plasma is available for delayed time over 4 hours.
 3. Frozen whole blood is not available.

Factor V (5)

CODE
SPECIMEN 2 mL Platelet Poor Citrated Plasma (1:9)
TEMPERATURE Frozen
SETUP SCHEDULE Contact Customer Care
TAT 14 days
METHODOLOGY Clot Based
PATIENT PREPARATION Not Applicable
COMMENT/S 1. The clotted or hemolytic specimens are not adequate.
 2. Only frozen poor-platelet plasma is available for delayed time over 4 hours.
 3. Frozen whole blood is not available.

Factor VII (7)

CODE
SPECIMEN 2 mL Platelet Poor Citrated Plasma (1:9)
TEMPERATURE Frozen
SETUP SCHEDULE Contact Customer Care
TAT 14 days
METHODOLOGY Clot Based



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PATIENT PREPARATION Not Applicable

COMMENT/S

1. The clotted or hemolytic specimens are not adequate.
2. Only frozen poor-platelet plasma is available for delayed time over 4 hours.
3. Frozen whole blood is not available.

Factor VIII (8)

CODE

SPECIMEN 2 mL Platelet Poor Citrated Plasma (1:9)

TEMPERATURE Frozen

SETUP SCHEDULE Contact Customer Care

TAT 14 days

METHODOLOGY Clot Based

PATIENT PREPARATION Not Applicable

COMMENT/S

1. The clotted or hemolytic specimens are not adequate.
2. Only frozen poor-platelet plasma is available for delayed time over 4 hours.
3. Frozen whole blood is not available.

Factor IX (9)

CODE

SPECIMEN 2 mL Platelet Poor Citrated Plasma (1:9)

TEMPERATURE Frozen

SETUP SCHEDULE Contact Customer Care

TAT 14 days

METHODOLOGY Clot Based

PATIENT PREPARATION Not Applicable

COMMENT/S

1. The clotted or hemolytic specimens are not adequate.
2. Only frozen poor-platelet plasma is available for delayed time over 4 hours.
3. Frozen whole blood is not available.

Factor X (10)

CODE

SPECIMEN 2 mL Platelet Poor Citrated Plasma (1:9)

TEMPERATURE Frozen

SETUP SCHEDULE Contact Customer Care

TAT 14 days

METHODOLOGY Clot Based

PATIENT PREPARATION Not Applicable

COMMENT/S

1. The clotted or hemolytic specimens are not adequate.
2. Only frozen poor-platelet plasma is available for delayed time over 4 hours.
3. Frozen whole blood is not available.

Factor XII (12)

CODE

SPECIMEN 2 mL Platelet Poor Citrated Plasma (1:9)

TEMPERATURE Frozen

SETUP SCHEDULE Contact Customer Care

TAT 14 days

METHODOLOGY Clot Based

PATIENT PREPARATION Not Applicable

COMMENT/S

1. The clotted or hemolytic specimens are not adequate.
2. Only frozen poor-platelet plasma is available for delayed time over 4 hours.
3. Frozen whole blood is not available.

Factor XIII (13)

CODE

SPECIMEN 2 mL Platelet Poor Citrated Plasma (1:9)

TEMPERATURE Frozen

SETUP SCHEDULE Contact Customer Care

TAT 14 days

METHODOLOGY Urea solubility test

PATIENT PREPARATION Not Applicable

COMMENT/S

1. The clotted or hemolytic specimens are not adequate.
2. Only frozen poor-platelet plasma is available for delayed time over 4 hours.
3. Frozen whole blood is not available.

Folate, RBC

CODE

SPECIMEN 4 mL EDTA Whole Blood

TEMPERATURE 2 – 8°C

SETUP SCHEDULE Contact Customer Care

TAT 14 days

METHODOLOGY ECLIA (Electrochemiluminescence Immunoassay)

PATIENT PREPARATION Not Applicable

COMMENT/S

Samples should not be taken from patients receiving therapy with high biotin doses (i.e. > 5 mg/day) until at least 8 hours following the last biotin administration.

It is contraindicated to measure samples of patients receiving therapy with certain pharmaceuticals, e.g. methotrexate or leucovorin, because of the cross-reactivity of folate binding protein with these compounds.

In rare cases, interference due to extremely high titers of antibodies to analyte-specific antibodies, streptavidin or ruthenium can occur.

Homocysteine

CODE

SPECIMEN 1 mL Serum

TEMPERATURE 2 – 8°C

SETUP SCHEDULE Contact Customer Care

TAT 14 days

METHODOLOGY High quality immunoassay

PATIENT PREPARATION Not Applicable

COMMENT/S

1. To minimize increases in homocysteine concentration from synthesis by red blood cells, place all specimens (serum and plasma) on ice after collection and prior to



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processing. Serum may clot more slowly and the volume of serum may be reduced as a result of being on ice.

2. Specimens from patients who are on drug therapy involving S-adenosyl-methionine may show falsely elevated levels of homocysteine. Specimens from patients taking methotrexate, carbamazepine, phenytoin, nitrous oxide, anticonvulsants or 6-azauridine triacetate may have elevated levels of homocysteine due to their effect on the metabolic pathway.

COMMENT/S

1. The clotted or hemolytic specimens are not adequate.
2. Only frozen poor-platelet plasma is available for delayed time over 4 hours.
3. Frozen whole blood is not available.

Immunoglobulin D

CODE

SPECIMEN 1 mL Serum

TEMPERATURE 2 – 8°C

SETUP SCHEDULE Contact Customer Care

TAT 14 days

METHODOLOGY TIA (Turbidimetric immunoassay)

PATIENT PREPARATION Not Applicable

COMMENT/S

Lupus Anticoagulant (Screen)

CODE

SPECIMEN 2 mL Platelet Poor Citrated Plasma (1:9)

TEMPERATURE Frozen

SETUP SCHEDULE Contact Customer Care

TAT 14 days

METHODOLOGY DRVVT (Dilute Russell's Viper Venom Time)

PATIENT PREPARATION Not Applicable

COMMENT/S

1. The clotted or hemolytic specimens are not adequate.
2. Only frozen poor-platelet plasma is available for delayed time over 4 hours.
3. Frozen whole blood is not available.

Indirect Coomb's Test

CODE

SPECIMEN 1 mL Serum

TEMPERATURE 2 – 8°C

SETUP SCHEDULE Contact Customer Care

TAT 14 days

METHODOLOGY Column agglutination method

PATIENT PREPARATION Not Applicable

COMMENT/S

Measles IgG

CODE

SPECIMEN 1 mL Serum

TEMPERATURE 2 – 8°C

SETUP SCHEDULE Contact Customer Care

TAT 14 days

METHODOLOGY CIA (Chemiluminescence immunoassay)

PATIENT PREPARATION Not Applicable

COMMENT/S

Inhibin-A

CODE

SPECIMEN 1 mL Serum

TEMPERATURE 2 – 8°C

SETUP SCHEDULE Contact Customer Care

TAT 14 days

METHODOLOGY CIA (Chemiluminescence immunoassay)

PATIENT PREPARATION Not Applicable

COMMENT/S Inhibin values fluctuate during the menstrual cycle. Inhibin levels in premenopausal women should be interpreted with caution.

Measles IgM

CODE

SPECIMEN 1 mL Serum

TEMPERATURE 2 – 8°C

SETUP SCHEDULE Contact Customer Care

TAT 14 days

METHODOLOGY CIA (Chemiluminescence immunoassay)

PATIENT PREPARATION Not Applicable

COMMENT/S

Lupus Anticoagulant (Confirm)

CODE

SPECIMEN 2 mL Platelet Poor Citrated Plasma (1:9)

TEMPERATURE Frozen

SETUP SCHEDULE Contact Customer Care

TAT 14 days

METHODOLOGY DRVVT (Dilute Russell's Viper Venom Time)

PATIENT PREPARATION Not Applicable

Procalcitonin

CODE

SPECIMEN 1 mL Serum

TEMPERATURE 2 – 8°C

SETUP SCHEDULE Contact Customer Care

TAT 14 days

METHODOLOGY CMIA (Chemiluminescence microparticle immunoassay)

PATIENT PREPARATION Not Applicable



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COMMENT/S	<p>Increased procalcitonin levels by non-bacterial causes are following:</p> <ul style="list-style-type: none"> - Neonates at < 48 hours of life (physiological elevation) - First day after a major trauma, major surgical intervention, severe burns, or treatment with OKT3 (muromonab-CD3) antibodies and other drugs stimulating the release of proinflammatory cytokines. - Patients with invasive fungal infections - Patients with acute attacks of Plasmodium falciparum malaria - Patients with prolonged or severe cardiogenic shock, prolonged severe organ perfusion anomalies, small cell lung cancer, severe liver cirrhosis and acute or chronic viral hepatitis, or medullary C-cell carcinoma of the thyroid <p>Specimens from patients who have received preparations of mouse monoclonal antibodies for diagnosis or therapy may contain human anti-mouse antibodies. Such specimens may show either falsely elevated or depressed values when tested with this assay. Heterophilic antibodies or rheumatoid factor in human serum can react with reagent immunoglobulins, interfering with in vitro immunoassays. Additional information may be required for diagnosis.</p>
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TAT	14 days
METHODOLOGY	TIA (Turbidimetric immunoassay)
PATIENT PREPARATION	Not Applicable
COMMENT/S	

Thyroid-Stimulating Immunoglobulin (TSI)

CODE	
SPECIMEN	1 mL Serum
TEMPERATURE	2 – 8°C
SETUP SCHEDULE	Contact Customer Care
TAT	14 days
METHODOLOGY	Bio assay
PATIENT PREPARATION	Not Applicable
COMMENT/S	

Protein S Ag, total

CODE	
SPECIMEN	1 mL Citrated Plasma (1:9)
TEMPERATURE	Frozen
SETUP SCHEDULE	Contact Customer Care
TAT	14 days
METHODOLOGY	ELISA (Enzyme linked immunosorbent assay)
PATIENT PREPARATION	Not Applicable
COMMENT/S	<ol style="list-style-type: none"> 1) Sodium citrated plasma should be removed immediately after venipuncture and stored frozen (-70°C). 2) Each coagulation assay requested should have its own vial. 3) If multiple coagulation assays are ordered, each test must be on a separate order.

Thyrotropin Receptor Ab, S

CODE	
SPECIMEN	1 mL Serum
TEMPERATURE	2 – 8°C
SETUP SCHEDULE	Contact Customer Care
TAT	14 days
METHODOLOGY	ECLIA (Electrochemiluminescence immunoassay)
PATIENT PREPARATION	Not Applicable
COMMENT/S	<p>This Thyrotropin Receptor Antibody test using quantitative electrochemiluminescence method is not to be confused with a Thyroid Stimulating Immunoglobulin test using bioassay.</p> <p>Eight hours before this blood test do not take multivitamins or dietary supplements containing biotin, which are commonly found in hair, skin, and nail supplements and multivitamins.</p> <p>Do not use samples from patients under sodium heparin treatment. Fractionated heparin (Clexane) showed interference at a concentration > 50 IU/L.</p> <p>In rare cases, interference due to extremely high titers of antibodies to test-specific antibodies, streptavidin or ruthenium can occur.</p>

Protein S Ag, Free

CODE	
SPECIMEN	1 mL Citrated Plasma (1:9)
TEMPERATURE	Frozen
SETUP SCHEDULE	Contact Customer Care
TAT	14 days
METHODOLOGY	ELISA (Enzyme linked immunosorbent assay)
PATIENT PREPARATION	Not Applicable
COMMENT/S	<ol style="list-style-type: none"> 1) Sodium citrated plasma should be removed immediately after venipuncture and stored frozen (-70°C). 2) Each coagulation assay requested should have its own vial. 3) If multiple coagulation assays are ordered, each test must be on a separate order.

Vitamin A (Retinol)

CODE	
SPECIMEN	2 mL Serum, Protected from Light
TEMPERATURE	Frozen
SETUP SCHEDULE	Contact Customer Care
TAT	14 days
METHODOLOGY	HPLC (High performance liquid chromatography)
PATIENT PREPARATION	Not Applicable
COMMENT/S	<ol style="list-style-type: none"> 1. Light should be blocked 2. After specimens collection, the specimen should be stored frozen. (-20°C or below)

Rheumatoid Factor (Qualitative)

CODE	
SPECIMEN	1 mL Serum
TEMPERATURE	2 – 8°C
SETUP SCHEDULE	Contact Customer Care

Vitamin B1 (Thiamine)

CODE	
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SPECIMEN	3 mL EDTA Whole Blood, Protected from Light
TEMPERATURE	Frozen
SETUP SCHEDULE	Contact Customer Care
TAT	14 days
METHODOLOGY	HPLC (High performance liquid chromatography)
PATIENT PREPARATION	Not Applicable
COMMENT/S	<ol style="list-style-type: none"> 1. Light should be blocked 2. After specimens collection, the specimen should be stored frozen.(-20°C or below)

Vitamin B2 (riboflavin)

CODE	
SPECIMEN	3 mL EDTA Whole Blood, Protected from Light
TEMPERATURE	Frozen
SETUP SCHEDULE	Contact Customer Care
TAT	14 days
METHODOLOGY	HPLC (High performance liquid chromatography)
PATIENT PREPARATION	Not Applicable
COMMENT/S	<ol style="list-style-type: none"> 1. Light should be blocked 2. After specimens collection, the specimen should be stored frozen.(-20°C or below)

Vitamin B2 (riboflavin)

CODE	
SPECIMEN	2 mL EDTA Plasma, Protected from Light
TEMPERATURE	Frozen
SETUP SCHEDULE	Contact Customer Care
TAT	14 days
METHODOLOGY	HPLC (High performance liquid chromatography)
PATIENT PREPARATION	Not Applicable
COMMENT/S	<ol style="list-style-type: none"> 1. Light should be blocked 2. After specimens collection, the specimen should be stored frozen.(-20°C or below)

Vitamin C (Ascorbic acid)

CODE	
SPECIMEN	2 mL Serum, Protected from Light
TEMPERATURE	Frozen
SETUP SCHEDULE	Contact Customer Care
TAT	14 days
METHODOLOGY	HPLC (High performance liquid chromatography)
PATIENT PREPARATION	Not Applicable
COMMENT/S	<ol style="list-style-type: none"> 1. Collection after 8 hours of NPO 2. Light should be blocked 3. After specimens collection, the specimen should be stored frozen.(-20°C or below)

Vitamin E (Tocopherol)

CODE	
SPECIMEN	2 mL Serum, Protected from Light
TEMPERATURE	Frozen
SETUP SCHEDULE	Contact Customer Care
TAT	14 days
METHODOLOGY	HPLC (High performance liquid chromatography)
PATIENT PREPARATION	Not Applicable
COMMENT/S	<ol style="list-style-type: none"> 1. Light should be blocked 2. After specimens collection, the specimen should be stored frozen.(-20°C or below)

Von Willebrand factor (vWF) activity (VWF:RCo, ristocetin cofactor)

CODE	
SPECIMEN	2 mL Citrated Plasma (1:9)
TEMPERATURE	Frozen
SETUP SCHEDULE	Contact Customer Care
TAT	14 days
METHODOLOGY	Fixation platelets and platelet agglutination test
PATIENT PREPARATION	Not Applicable
COMMENT/S	<ol style="list-style-type: none"> 1) Sodium citrated plasma should be removed immediately after venipuncture and stored frozen (-70°C). 2) Each coagulation assay requested should have its own vial. 3) If multiple coagulation assays are ordered, each test must be on a separate order.

Von Willebrand factor (vWF) Ab (VWF:Ag, VIII-related Ag)

CODE	
SPECIMEN	2 mL Citrated Plasma (1:9)
TEMPERATURE	Frozen
SETUP SCHEDULE	Contact Customer Care
TAT	14 days
METHODOLOGY	ELISA (Enzyme linked immunosorbent assay)
PATIENT PREPARATION	Not Applicable
COMMENT/S	<ol style="list-style-type: none"> 1) Sodium citrated plasma should be removed immediately after venipuncture and stored frozen (-70°C). 2) Each coagulation assay requested should have its own vial. 3) If multiple coagulation assays are ordered, each test must be on a separate order.